



Marine Biodiversity Insights from Indonesia: Cryptic Species and Chemical Profiles in the Nudibranch Genus *Chromodoris*

Muhammad Rizki Ramadhan^{1,*} and Sofyatuddin Karina¹

¹ Department of Marine Science, Faculty of Marine and Fisheries, Universitas Syiah Kuala, Banda Aceh 23111, Indonesia

1 Abstract

2 During a biodiversity survey on Weh Island, 25
3 Indonesia, marine gastropods of the genus 26
4 *Chromodoris* were investigated. Phylogenetic 27
5 analysis of 82 specimens revealed cryptic diversity 28
6 within *Chromodoris lochi*, indicating hidden genetic 29
7 lineages. Their prey-specific sponge (*Spongian* 30
8 sp.) was also collected to evaluate chemical 31
9 composition. Spectroscopic analysis identified 32
10 scalarane diterpenoids in both nudibranchs 33
11 and sponges, suggesting dietary sequestration or shared 34
12 biosynthetic origins. Molecular networking further 35
13 showed that chemical profiles of *Chromodoris* 36
14 were partially mixed among different clades, 37
15 contributing to apparent metabolite variation. 38
16 Descriptive statistical analysis demonstrated higher 39
17 concentrations of biochemical compounds in 40
18 specimens from northern compared to southern Weh 41
19 Island. These findings emphasize the integration 42
20 of phylogenetics and metabolomics to understand 43
21 spatial and cryptic diversity in marine chemical 44
22 ecology.

23 **Keywords:** Biodiversity, *Chromodoris*, Molecular Network,
24 Weh Island.

1 Introduction

25 *Chromodoris* is one of the largest genera of brightly 26 colored nudibranchs distributed throughout tropical 27 and subtropical Indo-Pacific regions. Molecular 28 phylogenetic studies have demonstrated that this 29 genus comprises genetically distinct lineages despite 30 morphological similarity among species [1,2]. 31 Members of *Chromodoris* commonly inhabit coral reef 32 ecosystems and display selective feeding behavior, 33 preying on specific sponge taxa. During feeding, these 34 nudibranchs sequester sponge-derived secondary 35 metabolites and accumulate them in mantle dermal 36 formations (MDFs), where the compounds are 37 modified and utilized for chemical defense [3]. This 38 process indicates the presence of detoxification and 39 transport mechanisms that enable the safe storage of 40 toxic metabolites.

41 Recent genome mining studies have revealed that 42 sponge-associated microorganisms play a crucial 43 role in the biosynthesis of secondary metabolites 44 [4]. These uncultured marine microbes, often 45 residing on sponge surfaces, may be co-ingested with 46 sponge tissue, thereby contributing microbial-derived 47 compounds to nudibranch chemical profiles. Several 48 sponge-microbe symbioses have been reported. 49 For example, *Lamellodysidea herbacea* and *Dysidea* 50 *granulosa* harbor the cyanobacterium *Synechococcus* 51 *elongatus*, which produces polybrominated diphenyl 52 ethers [5]. Similarly, *Theonella swinhonis* contains 53 polyketide-producing *Pseudomonas* species [6], while

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*Corresponding author:
Muhammad Rizki Ramadhan
muhammadrizkiramadhan1@outlook.com

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55 *Acanthostrongylophora ingens* hosts *Micromonospora* sp.,
56 known to biosynthesize manzamine A [7].

57 Chemical investigations of nudibranchs have
58 consistently demonstrated prey-specific metabolite
59 patterns. For instance, *C. luteorosea* accumulates
60 diterpenoids from *Dendrilla* sp., *Dysidea* sp.,
61 *Chelonaplysilla* sp., and *Aplysilla polyrhaphis* [8].
62 Likewise, *C. elisabethina* and *C. magnifica* prey on
63 *Heteronema* sp., which contains puerphenone, whereas
64 *C. lochi* feeds on *Luffariella variabilis*, a known source of
65 manoalide [9]. During our recent expedition around
66 Weh Island, *Chromodoris* specimens were found to
67 contain scalarane-type diterpenoids, compounds
68 recognized for antibacterial, antifungal, antifouling,
69 anticancer, and pheromonal activities [10–12].

70 Despite increasing evidence of sponge-derived
71 metabolites in nudibranchs, the ecological and
72 chemical relationships among nudibranchs, sponge
73 prey, and associated microbes remain insufficiently
74 understood. Therefore, this study aims to elucidate the
75 biological diversity of *Chromodoris* and characterize the
76 chemical profiles of both nudibranchs and associated
77 uncultured sponge microbes using molecular
78 networking approaches.

79 2 Methodology

80 2.1 General

81 NMR spectra were recorded on a Bruker 600 MHz
82 spectrometer using deuterated chloroform (CDCl_3 ,
83 Cambridge Isotope Laboratories) as solvent at ambient
84 temperature. Tetramethylsilane (TMS) was used as
85 the internal reference standard. Spectral acquisition
86 and data processing were performed using TopSpin
87 4.0 (Bruker BioSpin).

88 High-resolution electrospray ionization mass
89 spectrometry (HRESIMS) analyses were conducted
90 on an Amazon Ion Trap mass spectrometer (Bruker
91 Daltonics, Bremen, Germany) coupled to an Agilent
92 1260 Infinity LC system (Agilent Technologies, Santa
93 Clara, CA, USA). Chromatographic separation was
94 achieved using a reversed-phase C18 analytical HPLC
95 column (Phenomenex, Torrance, CA, USA; 5 m, 250
96 \times 4.6 mm). Mass spectral data were processed and
97 interpreted using Compass DataAnalysis 4.2 (Bruker
98 Daltonics).

99 2.2 Biomaterial

100 The cryptic species of *Chromodoris lochi* and its
101 prey sponges were collected around Weh Island in
102 2018. *Chromodoris lochi* and sponge species have

been identified and deposited by our collaborator at
Naturalis Biodiversity Center, Leiden, the Netherlands.

103 2.3 Isolation and Identification

106 Fresh specimens were immediately extracted
107 with acetone at room temperature to prevent
108 metabolite degradation. The crude extracts
109 were concentrated under reduced pressure and
110 subsequently partitioned between dichloromethane
111 and water to afford a lipophilic organic fraction. The
112 dichloromethane extract was subjected to silica gel
113 column chromatography using a stepwise gradient of
114 n-hexane–dichloromethane to obtain the non-polar
115 fraction. The resulting fractions were analyzed
116 by LC–MS/MS to generate fragmentation data for
117 molecular networking analysis. MS/MS features were
118 processed and organized to construct a molecular
119 network, enabling visualization of chemical diversity
120 and metabolite distribution among *Chromodoris*
121 nudibranch specimens.

122 2.4 Feature-Based Molecular Networking

123 MS/MS data were processed using MZmine 2 for
124 peak detection, deconvolution, alignment, and
125 feature grouping to reconstruct the molecular
126 network. Following data processing, a quantitative
127 feature table was exported in CSV format, and
128 MS/MS spectra for each feature were generated in
129 MGF format. The raw MS/MS dataset has been
130 deposited in the MassIVE repository under accession
131 number MSV000087358 (<https://massive.ucsd.edu>).
132 For molecular networking, precursor ion mass
133 tolerance and fragment ion tolerance were set to
134 0.05 Da. Network construction parameters included
135 a minimum of 10 matched fragment peaks and a
136 cosine similarity score threshold of 0.6. The processed
137 data were uploaded to the Global Natural Products
138 Social Molecular Networking (GNPS) platform
139 (<https://gnps.ucsd.edu>) for network generation. The
140 resulting molecular networks were visualized and
141 analyzed using Cytoscape version 3.7.2.

142 3 Results

143 3.1 Identification of Cryptic species

144 Molecular phylogenetic analysis of 70 *Chromodoris*
145 *lochi* specimens revealed the presence of cryptic
146 lineages, supporting hidden genetic diversity within
147 this morphospecies (Figure 1). In contrast, all prey
148 sponges associated with *C. lochi* were taxonomically
149 identified as *Spongian* sp. Chemical characterization
150 of the crude extracts was performed using ^1H



151 NMR spectroscopy and MS/MS analysis. These
 152 analyses led to the identification of well-known
 153 diterpenoid and sesterterpenoid metabolites,
 154 consistent with previously reported compounds in
 155 related nudibranch–sponge systems. Furthermore,
 156 feature-based molecular networking enabled
 157 visualization of metabolite distribution patterns, while
 158 environmental correlation analysis provided insight
 159 into spatial variations in chemical composition.

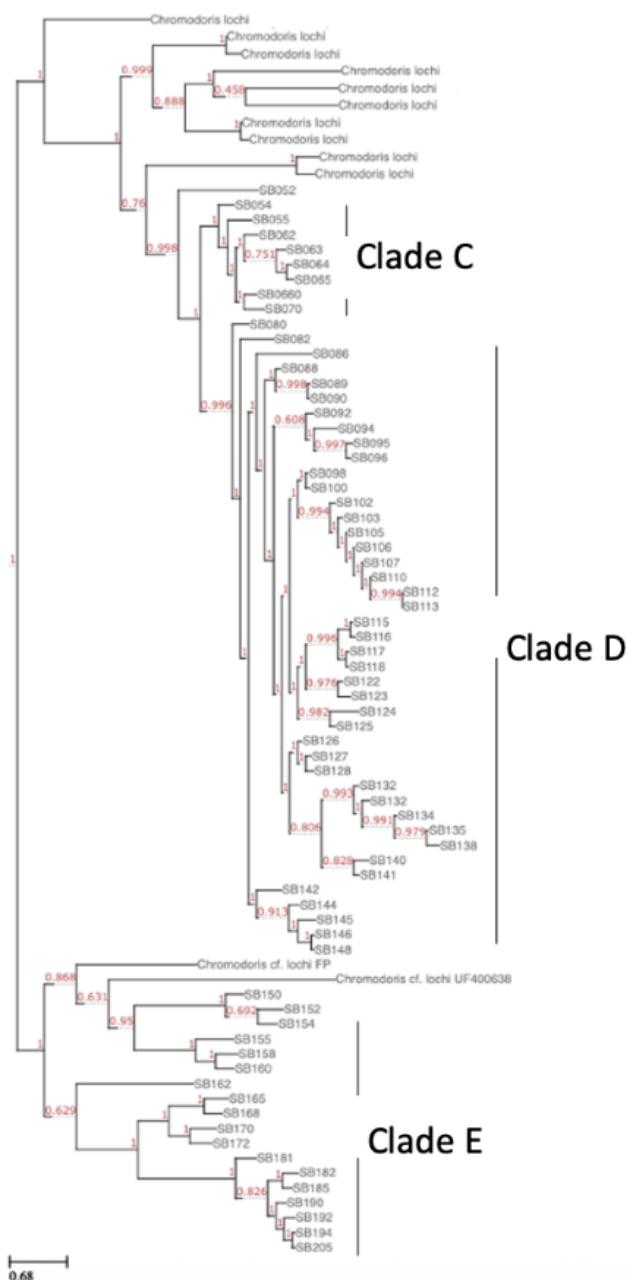


Figure 1. Molecular phylogenetic of *Chromodoris* nudibranchs

3.2 Chemical Profiling

All lipophilic extracts were identified as well-known scalarane diterpenoids by comparison of their ¹H NMR and HRESIMS data with previously reported data (Gonzalez, 2010). The presence of chemical diversity in the extracts suggests that common chemical reactions, such as oxidation, cyclization, Michael addition, and Diels–Alder reactions, may be involved in enzymatic processes within the nudibranch (Figure 2).

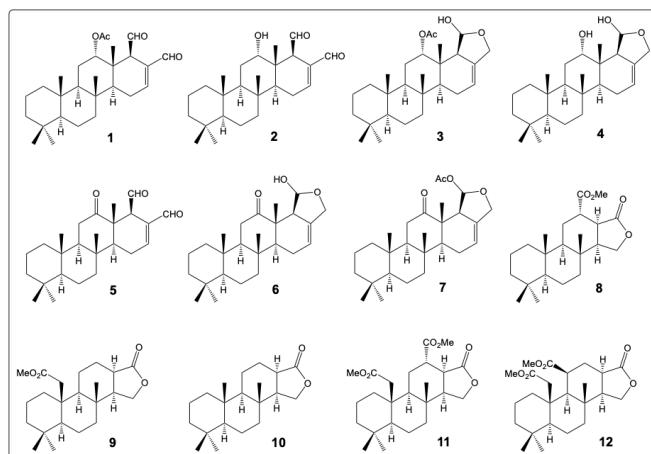


Figure 2. Scalarane diterpenoids from *Chromodoris* nudibranchs

3.3 Metabolomic Profiling by MS/MS and Molecular Networking

The lipophilic extracts of *C. lochi*, together with reference compounds (1–12), were subjected to MS/MS analysis to comprehensively characterize their secondary metabolite profiles. Features were subsequently organized according to phylogenetic clades to assess chemotaxonomic patterns. Notably, clade C exhibited a distinct and more chemically diverse metabolomic signature compared to clades D and F, indicating potential lineage-specific metabolic differentiation.

For molecular network construction, raw MS/MS datasets were processed using MZmine2, yielding 1,852 and 1,624 aligned features, respectively (Figure 3). The resulting quantitative feature table (CSV format) was curated and imported into MetaboAnalyst for multivariate statistical evaluation. Principal component analysis (PCA) revealed a clear clustering pattern, with clade C distinctly separated from clades D and E along the principal components (Figure 4). In contrast, clades D and E showed considerable overlap, suggesting high chemical similarity between these groups.

194 The observed clustering pattern supports the
 195 hypothesis that cryptic species within clades D
 196 and E may share closely related biosynthetic
 197 capacities, whereas clade C likely undergoes divergent
 198 metabolic regulation. These findings highlight the
 199 relevance of integrating molecular phylogenetics with
 200 metabolomics to resolve chemotype differentiation
 201 within morphologically similar taxa.

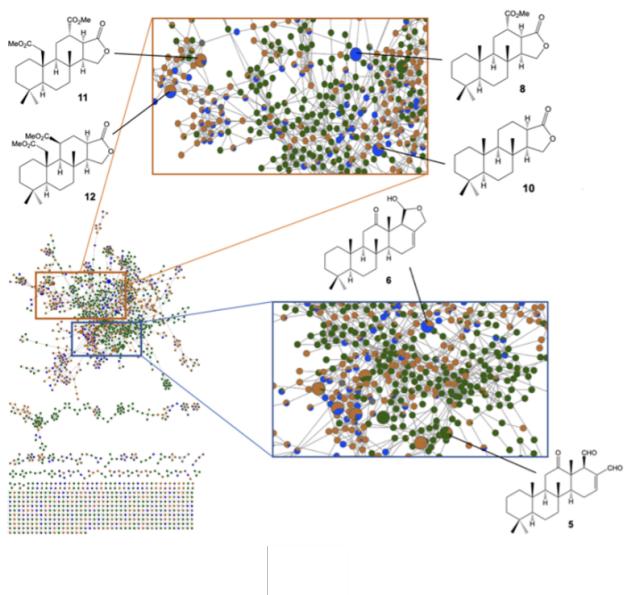


Figure 3. Feature-based molecular networking of *Chromodoris* nudibranchs.

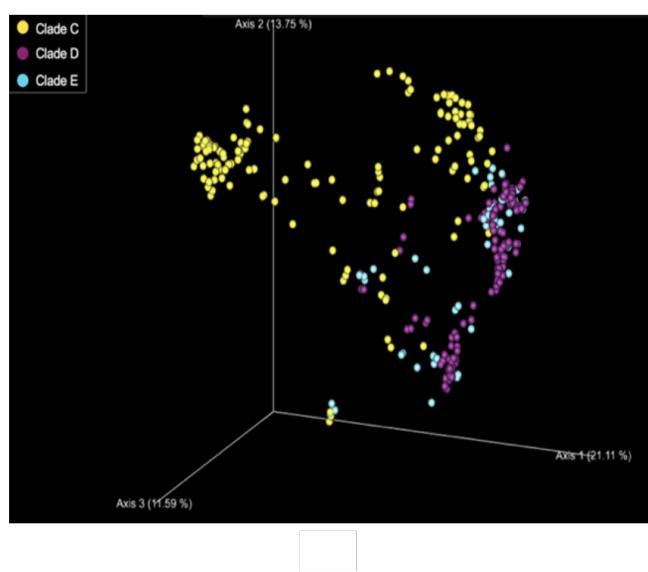


Figure 4. Principal Component Analysis result.

202 4 Discussion

203 The present study provides robust evidence for cryptic
 204 speciation within the nudibranch *Chromodoris lochi*,
 205 supported by molecular phylogenetic analyses of 82

206 specimens. These findings contribute to the growing
 207 recognition that morphology-based taxonomy
 208 may substantially underestimate species diversity,
 209 particularly among soft-bodied marine invertebrates
 210 such as nudibranchs. Cryptic species, genetically
 211 distinct yet morphologically indistinguishable,
 212 underscore the necessity of molecular approaches for
 213 accurate taxonomic resolution [12–13].

214 Phylogenetic reconstruction revealed multiple
 215 well-supported clades within what has historically
 216 been treated as a single species (Figure 1). The
 217 identification of distinct lineages (clades C, D, and
 218 E) indicates substantial evolutionary divergence
 219 despite apparent morphological stasis. Such patterns
 220 are consistent with both allopatric and sympatric
 221 speciation processes in marine systems, where
 222 ecological specialization or chemically mediated
 223 selection pressures may promote divergence even
 224 in the absence of obvious geographic barriers.
 225 Ecological observations demonstrated that all *C.*
 226 *lochi* specimens preyed upon the same sponge
 227 taxon (*Spongian* sp.), indicating a conserved trophic
 228 association across clades [13]. This dietary uniformity
 229 suggests that genetic divergence has occurred without
 230 detectable shifts in primary prey selection. Given
 231 the highly specialized sponge-feeding behavior
 232 characteristic of *Chromodoris*, selective retention of
 233 bioactive, chemically defended prey likely confers
 234 both nutritional and defensive advantages, reinforcing
 235 trophic conservatism across lineages [10].

236 Metabolomic profiling provided additional resolution
 237 into intra- and interclade chemical variation. Based on
 238 1H NMR and high-resolution MS/MS analyses, they
 239 identified a suite of scalarane diterpenoids and related
 240 terpenoid metabolites, consistent with previous
 241 reports of sponge-derived compounds sequestered
 242 by nudibranch tissues. These metabolites are not
 243 biosynthesized de novo but are acquired through
 244 dietary sequestration and may undergo subsequent
 245 enzymatic modification. Such trophically mediated
 246 chemical acquisition is a defining feature of the genus
 247 and highlights the central role of chemical ecology in
 248 nudibranch diversification [13].

249 Comprehensive MS/MS analysis combined with
 250 molecular networking (MZmine2 and MetaboAnalyst
 251 workflows) detected over 1,800 aligned molecular
 252 features. Principal Component Analysis (PCA)
 253 revealed that clade C forms a chemically distinct
 254 cluster relative to clades D and E, whereas clades
 255 D and E exhibit substantial overlap in metabolomic



space. This pattern suggests that some cryptic lineages diverge at both genetic and chemical levels (e.g., clade C), while others retain conserved metabolomic profiles despite clear genetic differentiation. The chemical similarity between clades D and E may reflect shared biosynthetic modification pathways, comparable prey-derived metabolite processing, or similar ecological constraints.

The evolutionary implications of these findings are multifaceted. First, distinct chemotypes among cryptic lineages support the use of metabolomic signatures as auxiliary taxonomic markers that complement phylogenetic data. Second, chemical divergence may act as a driver or consequence of speciation, potentially influencing ecological adaptation, predator deterrence efficiency, or reproductive signaling. Conversely, the persistence of conserved metabolite profiles across genetically distinct clades may indicate stabilizing selection maintaining functional chemical defenses.

Collectively, this study integrates phylogenetic, ecological, and metabolomic evidence to reveal previously unrecognized diversity within *C. lochi*. The coexistence of genetic divergence, trophic conservatism, and partial chemical differentiation underscores the complexity of nudibranch biodiversity and highlights the value of integrative approaches in marine systematics. These findings open new perspectives on the evolutionary mechanisms shaping chemical diversity in marine organisms.

5 Conclusion

This study provides compelling evidence for the existence of cryptic species within the nominal taxon *Chromodoris lochi*, underscoring the limitations of morphology-based taxonomy in accurately resolving biodiversity among marine nudibranchs. Molecular phylogenetic analyses revealed multiple well-supported genetic clades that likely represent distinct evolutionary lineages, despite their shared external morphology.

Notably, these genetically divergent clades maintain a conserved ecological niche, particularly in their specialized feeding on *Spongian* sp. sponges. Such trophic conservatism suggests that ecological stasis may persist despite genetic divergence, potentially driven by strong stabilizing selection associated with dietary specialization and chemically mediated defense strategies.

Metabolomic profiling further revealed both shared and clade-specific chemical signatures. While certain

clades displayed distinct chemotypes, others retained overlapping metabolomic profiles, indicating that chemical divergence does not uniformly accompany genetic differentiation. These findings highlight the complex interplay between phylogeny, ecology, and chemical acquisition in shaping nudibranch diversity. Moreover, the application of MS/MS-based molecular networking combined with multivariate statistical analyses proved highly effective in resolving chemodiversity patterns and supporting taxonomic inference.

Collectively, the integration of phylogenetic, ecological, and metabolomic datasets provides a multidimensional framework for understanding species boundaries in marine systems. This study reinforces the importance of integrative taxonomy and advances our understanding of the evolutionary processes underlying cryptic speciation and chemical diversification in nudibranchs.

Data Availability Statement

Data will be made available on request.

Author Contributions

M.R.R. and S.K. conceptualized and designed the study. M.R.R. conducted the experimental work, including methodology development, data analysis, data curation, and preparation of the original draft. Validation was carried out collaboratively by M.R.R. and S.K. All authors have reviewed and approved the final version of the manuscript and consent to its publication.

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353 Conflicts of Interest

354 The authors declare no conflicts of interest.

355 Ethical Approval and Consent to Participate

356 Not applicable.

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